

“Biological systems are necessarily metastable. They are created, modulated, and destroyed according to a temporal plan that meets the survival needs of the cell, organism, and species...Clearly, no biological system is close to true equilibrium or it would be dead...

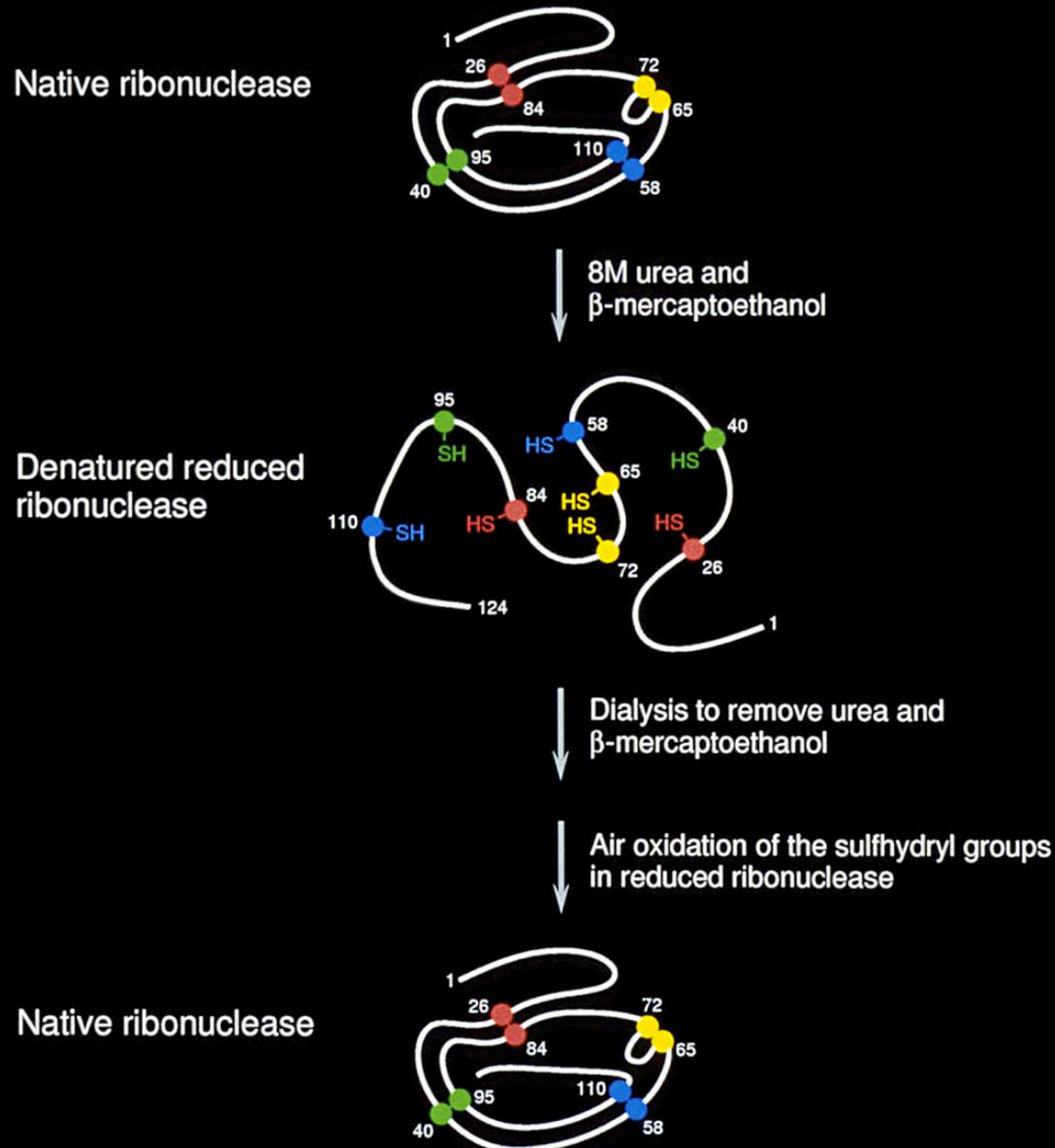
Systems survive by consuming free energy and regulating rates...a human turns over 40 kg of ATP daily, and 90% of the information in a genome encodes biological catalysts...

The more quickly something is done, the more difficult it is to do it accurately. Accuracy and specificity often are sacrificed in the name of speed...This balance is especially important in optimizing the rate of information transfer from the gene to the structures of functional nucleic acids and proteins.

The entropic activation barrier to prompt and accurate genetic expression...is overcome by the lavish use of nucleotide triphosphate to proofread and edit...”

Paul Sigler et al, 1998.

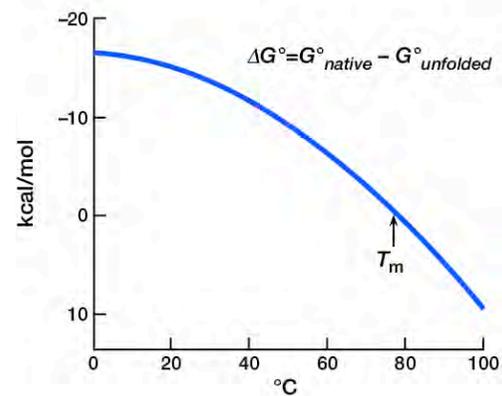
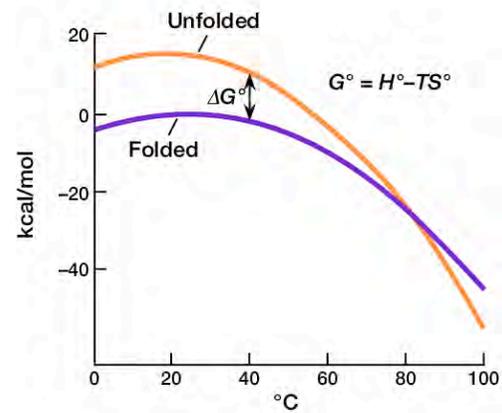
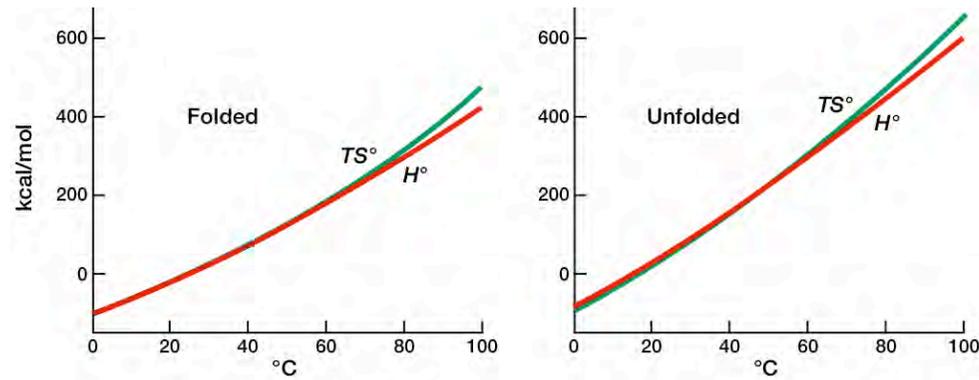
The amino acid sequence of a protein contains all of the information for folding into the unique native state, presumed to lie at the energetic minimum



Anfinsen et al, 1961

Thermodynamic Measurements

Enthalpic (H) and entropic (TS) contributions to the free energies of the folded and unfolded states of the enzyme lysozyme as a function of temperature—Large changes over T but net stability is only a few kcal/mole



Pfeil and Privalov, 1976,
adapted from Creighton text, 1997

Major Forces Contributing to the Conformational Stability of RNase T1

Stabilizing

Hydrogen bonding (104 HB) = 166 kcal/mole

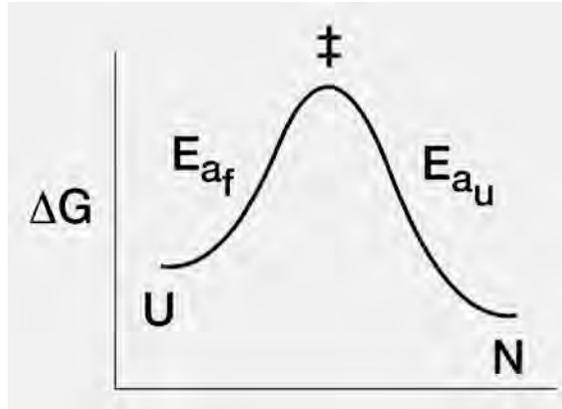
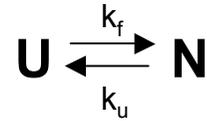
Hydrophobic groups buried
(entropy of dehydration) = 94

Destabilizing

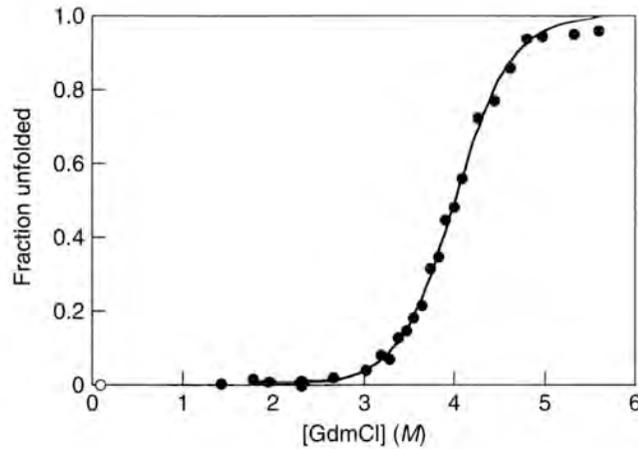
Conformational entropy = 177 kcal/mole

Peptide groups buried = 81

Behavior of Two-state Proteins, Typically Fewer Than 100 Amino Acids



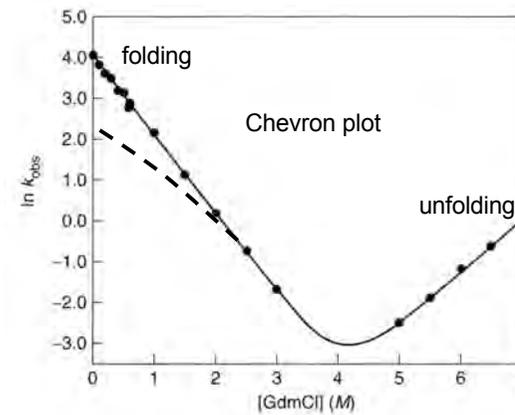
Equilibrium studies



Cooperative transition

$$\Delta G_{(U \rightarrow N)} = -RT \ln K_{eq}$$

Kinetic studies

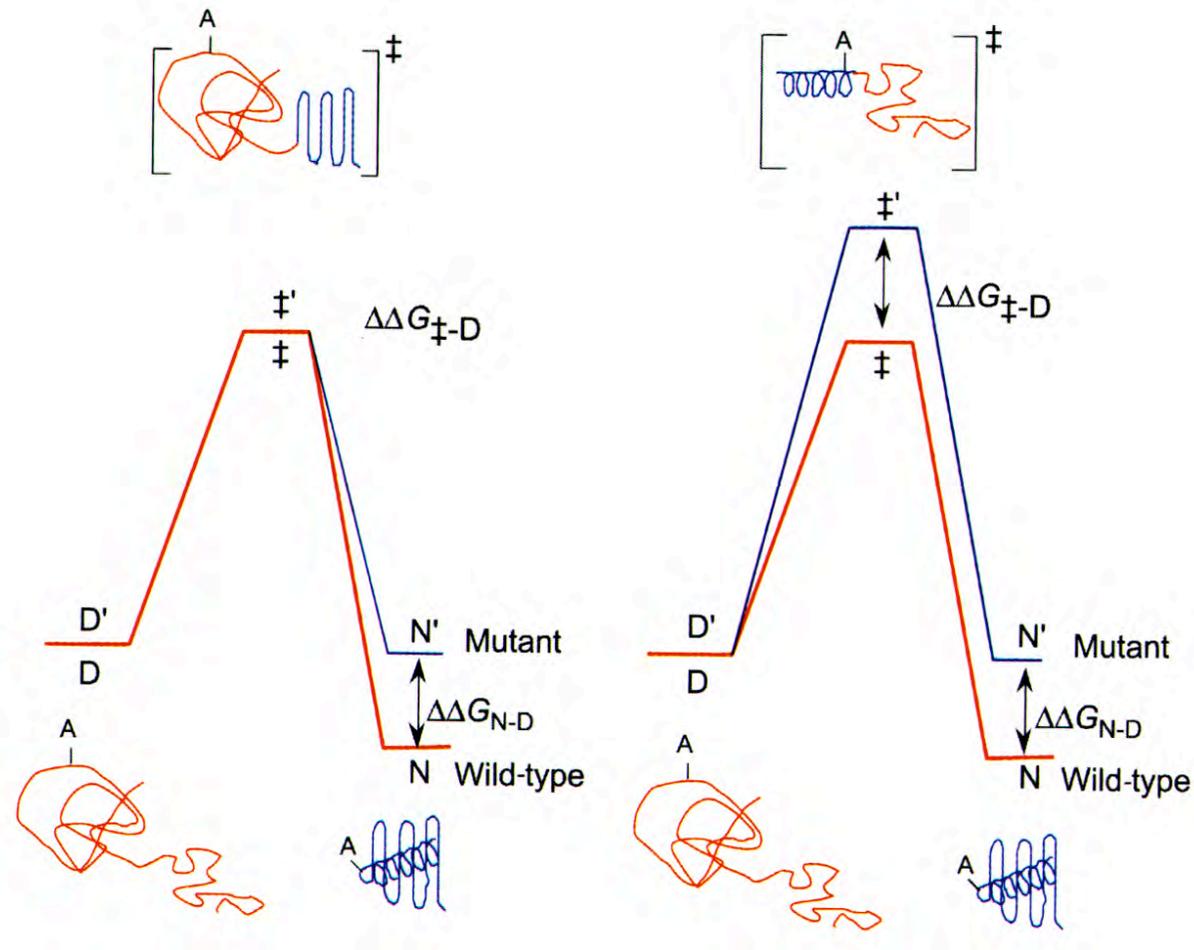


$$\ln k_{obs} = \ln ((k_f^{H_2O}) \exp(-m_{k_f} [\text{denaturant}]) + k_u^{H_2O} \exp(m_{k_u} [\text{denaturant}]))$$

E_a $RT \ln k$ (but non-Arrhenius behavior)

Deviation from “V” indicates formation of an intermediate or multimolecular aggregation

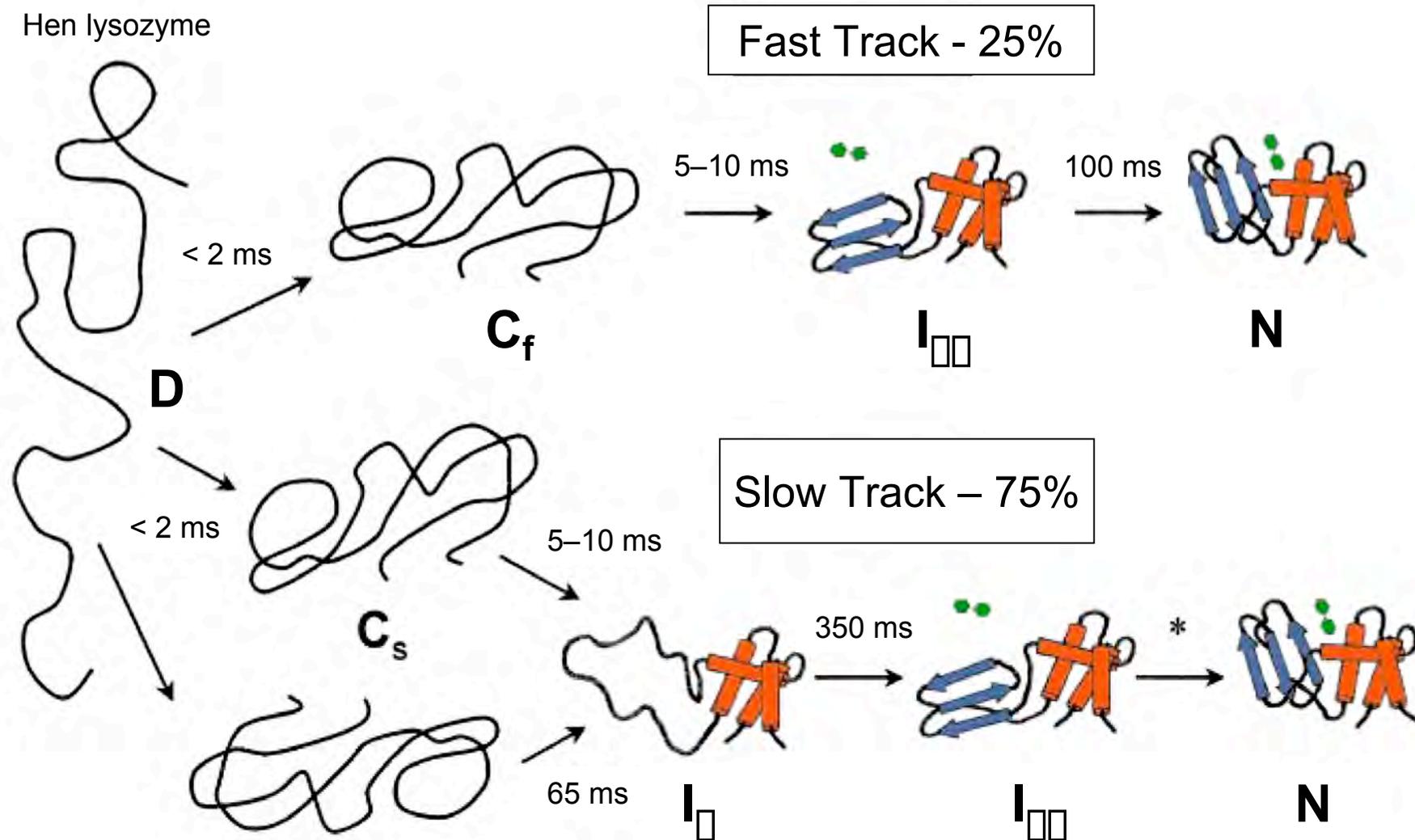
Deriving structure of transition state of folding for 2-state proteins by mutational analysis by comparing rates of folding and unfolding of wt and mutant



from Fersht and Daggett, 2002

Larger proteins fold in a more complex manner, involving intermediates and multiple routes

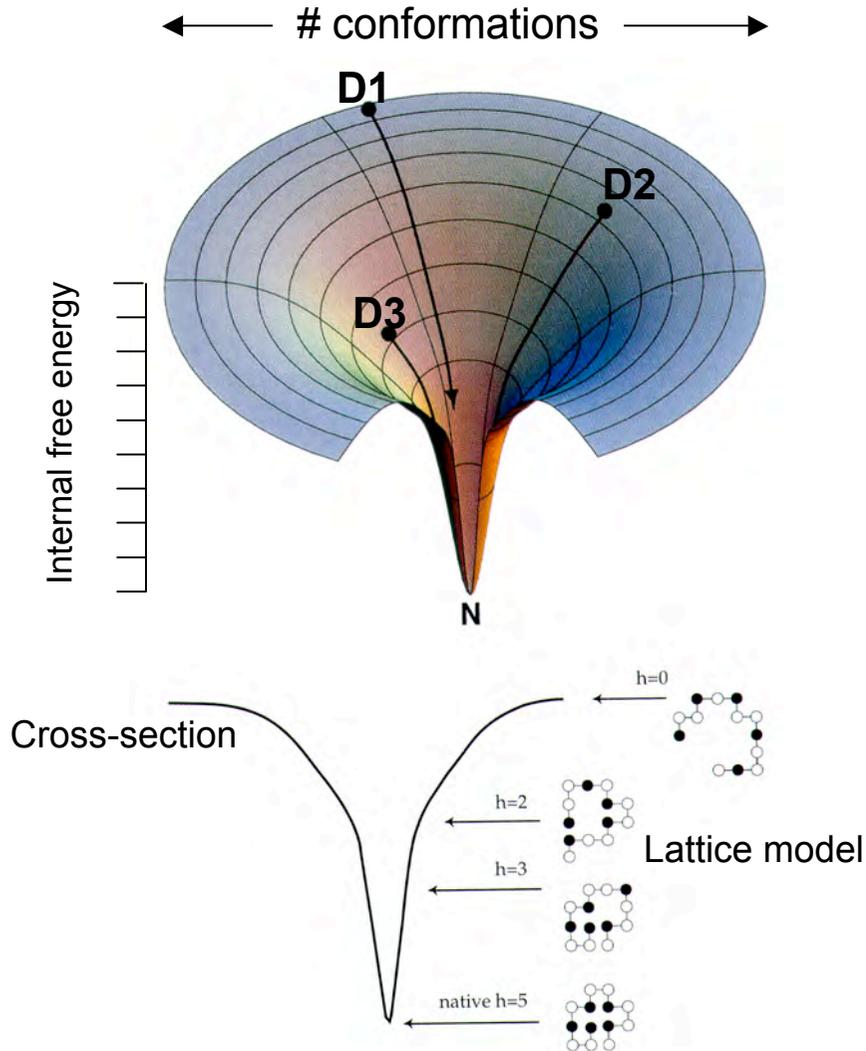
Hen lysozyme



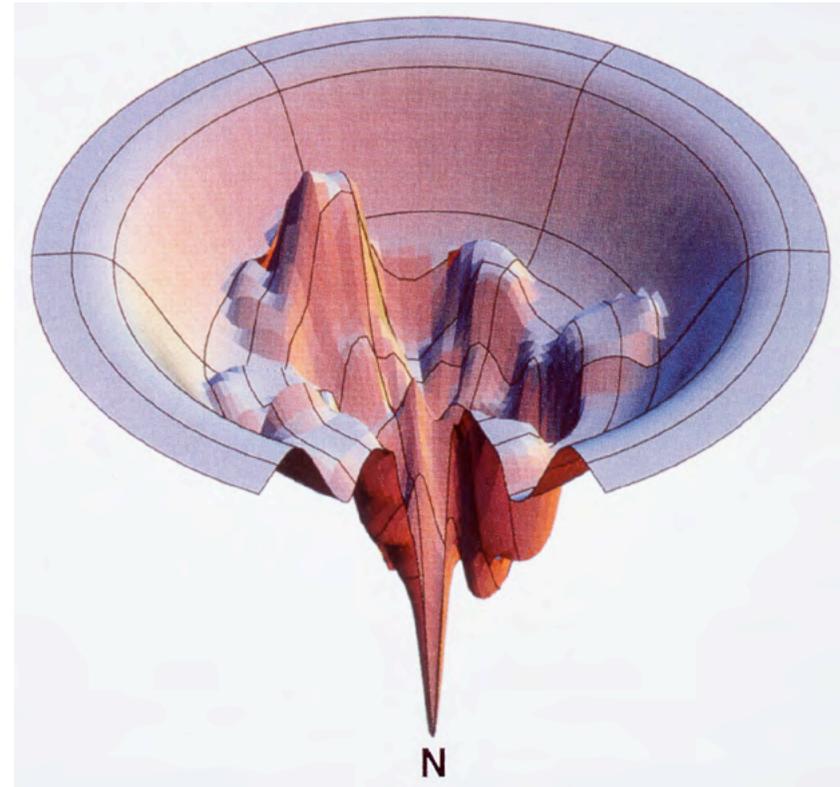
From Matagne, Dobson, & coworkers, 2000

Energy “landscapes” connect single-polypeptide microscopics with experimental macroscopics

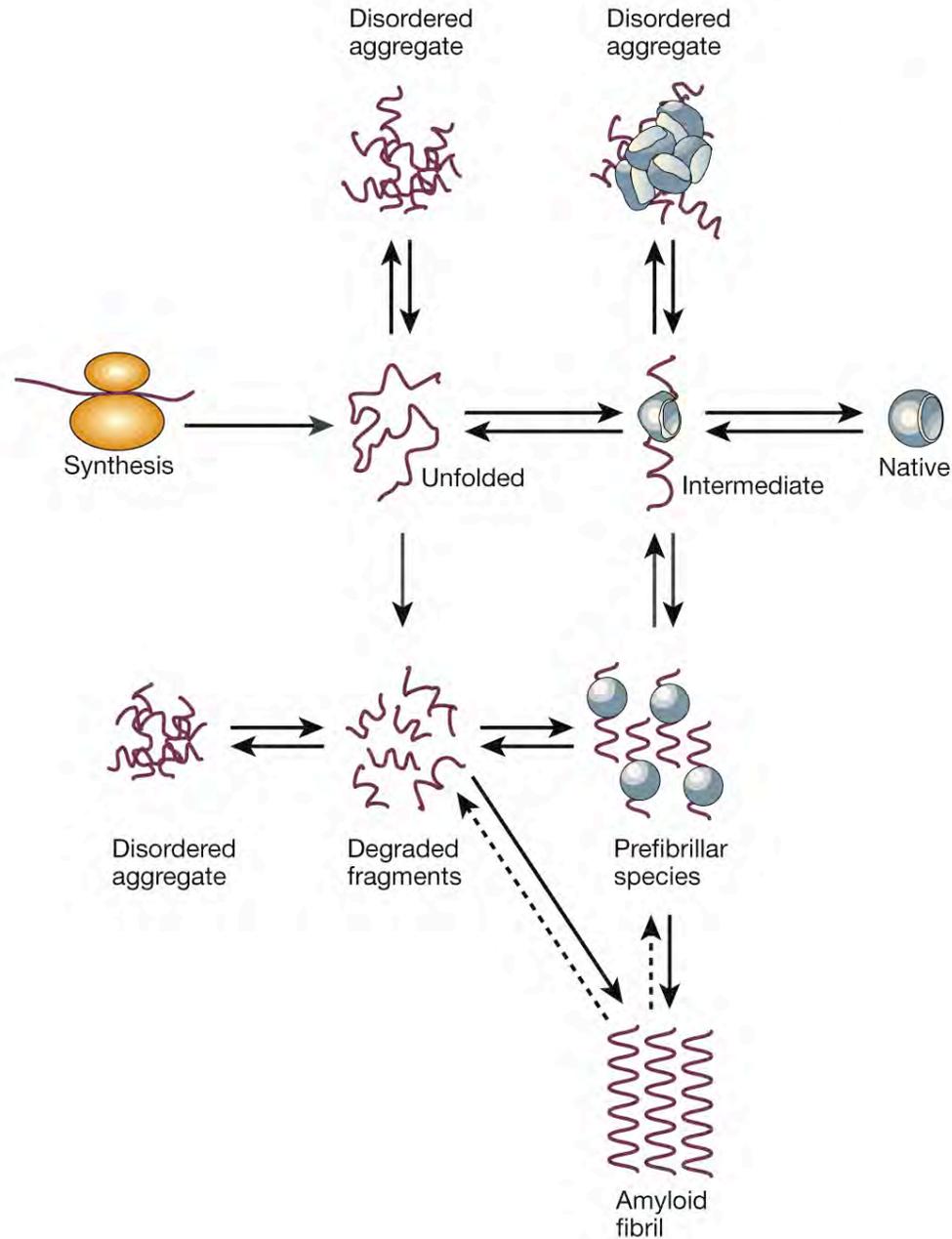
Two-state Landscape



Rugged landscape replete with kinetic “traps”



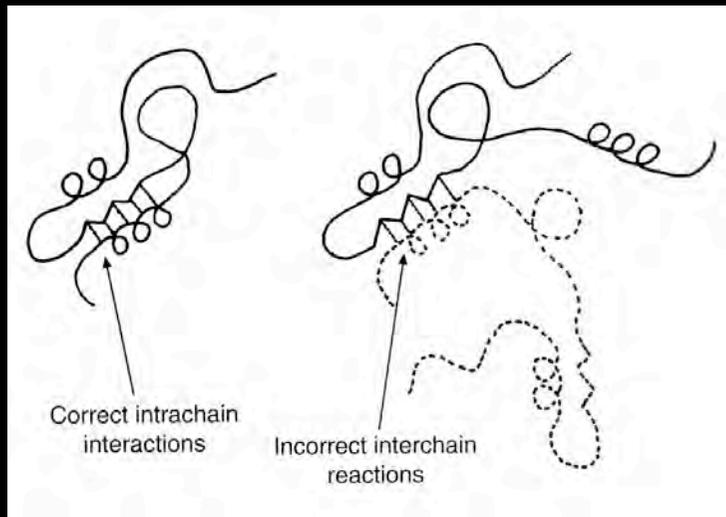
Fate of misfolded polypeptide chains in the cell – multimolecular “disordered” aggregates and, in ~20 cases, “ordered” amyloid fibrils



Adapted from Dobson, 2003

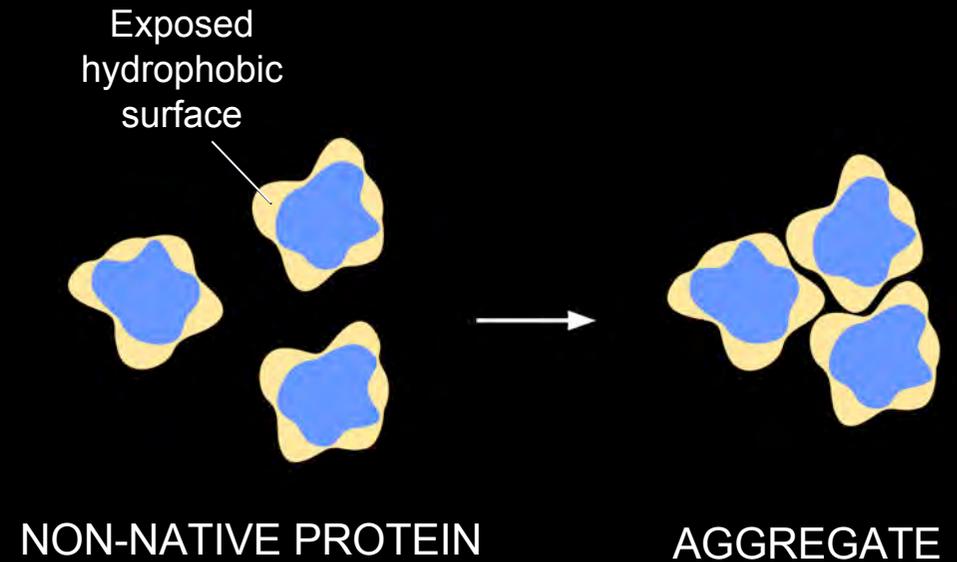
Mechanisms of Protein Aggregation

Domain Swapping



M.E. Goldberg and coworkers, 1974

Hydrophobic Interactions



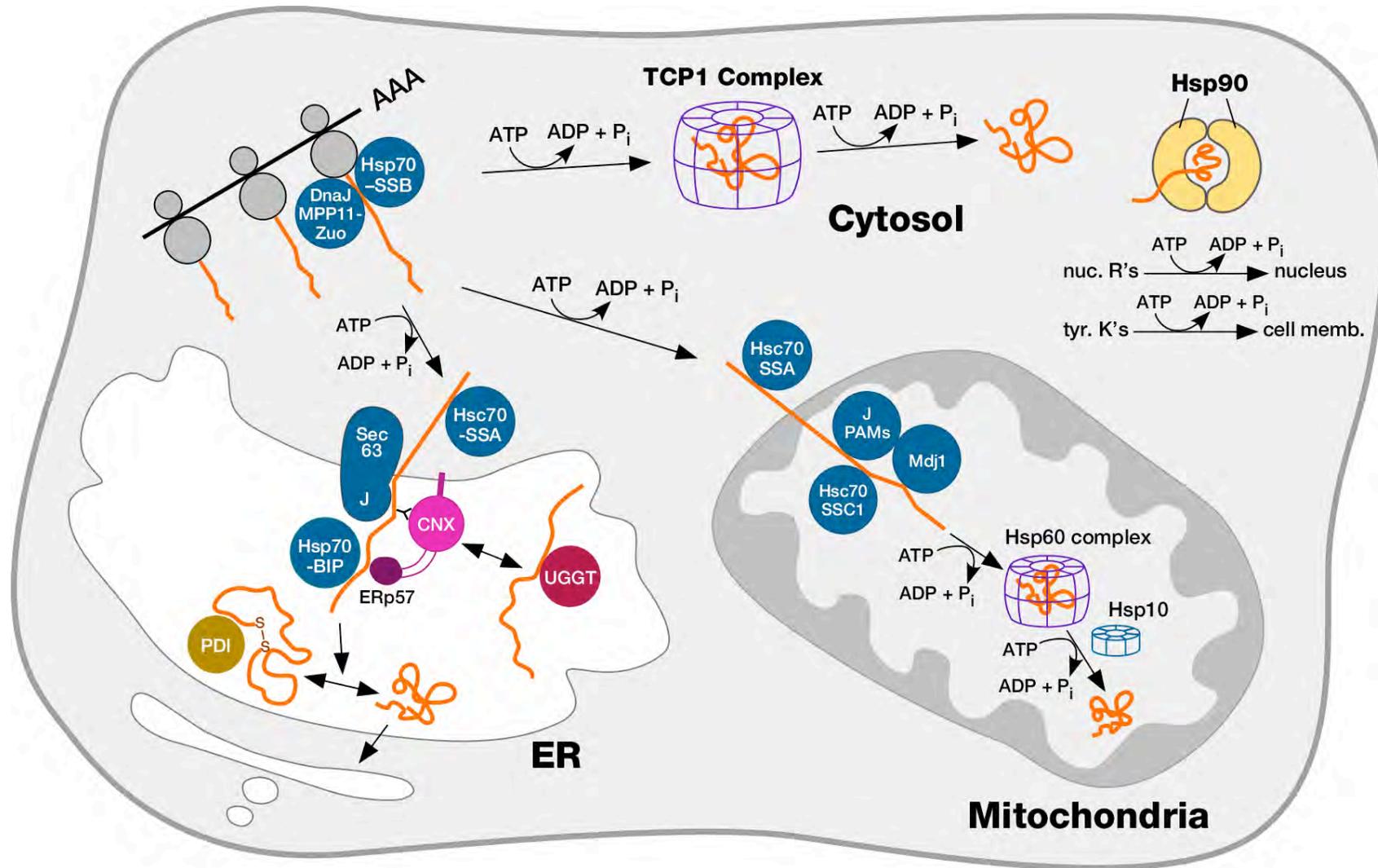
NATIVE PROTEIN

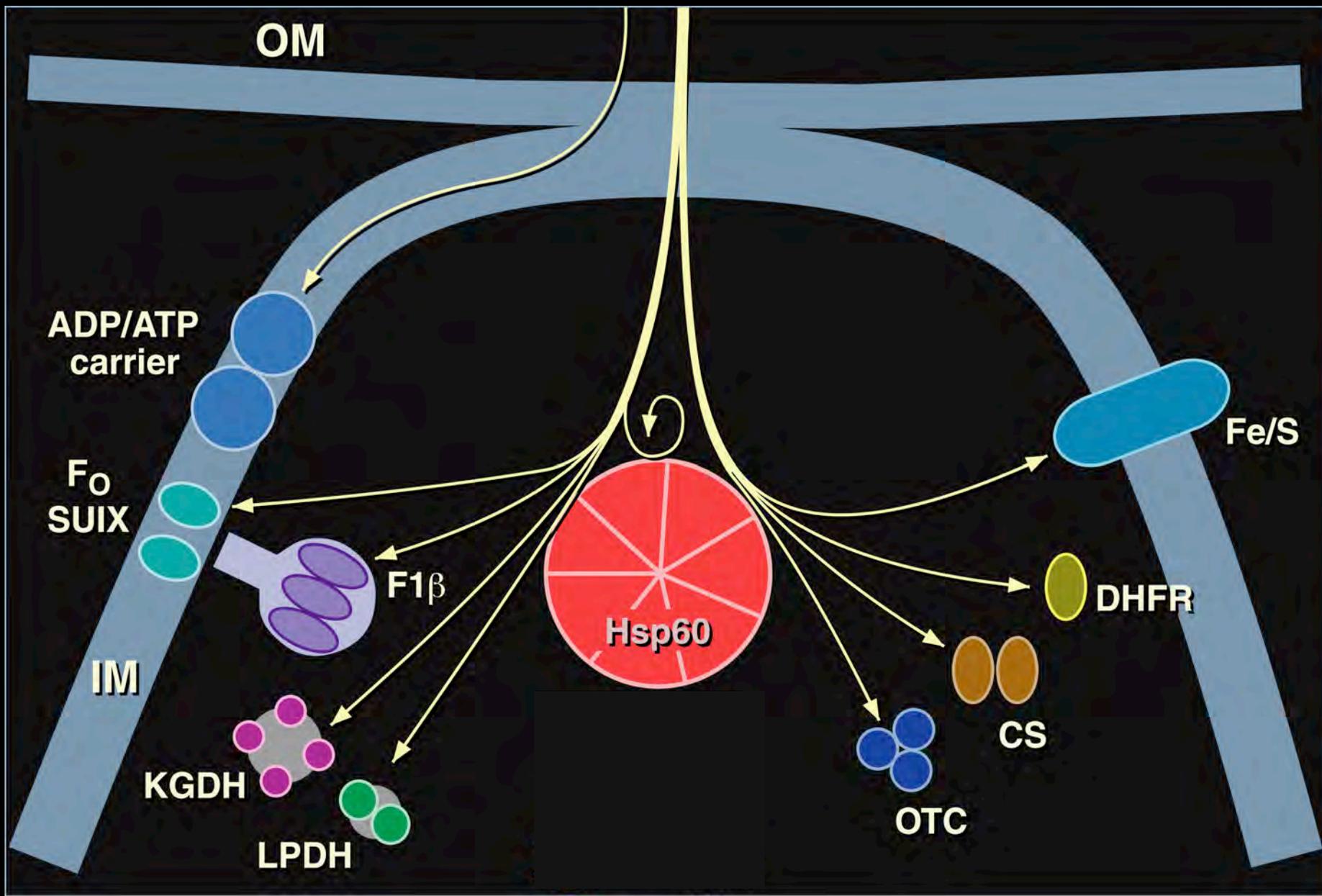


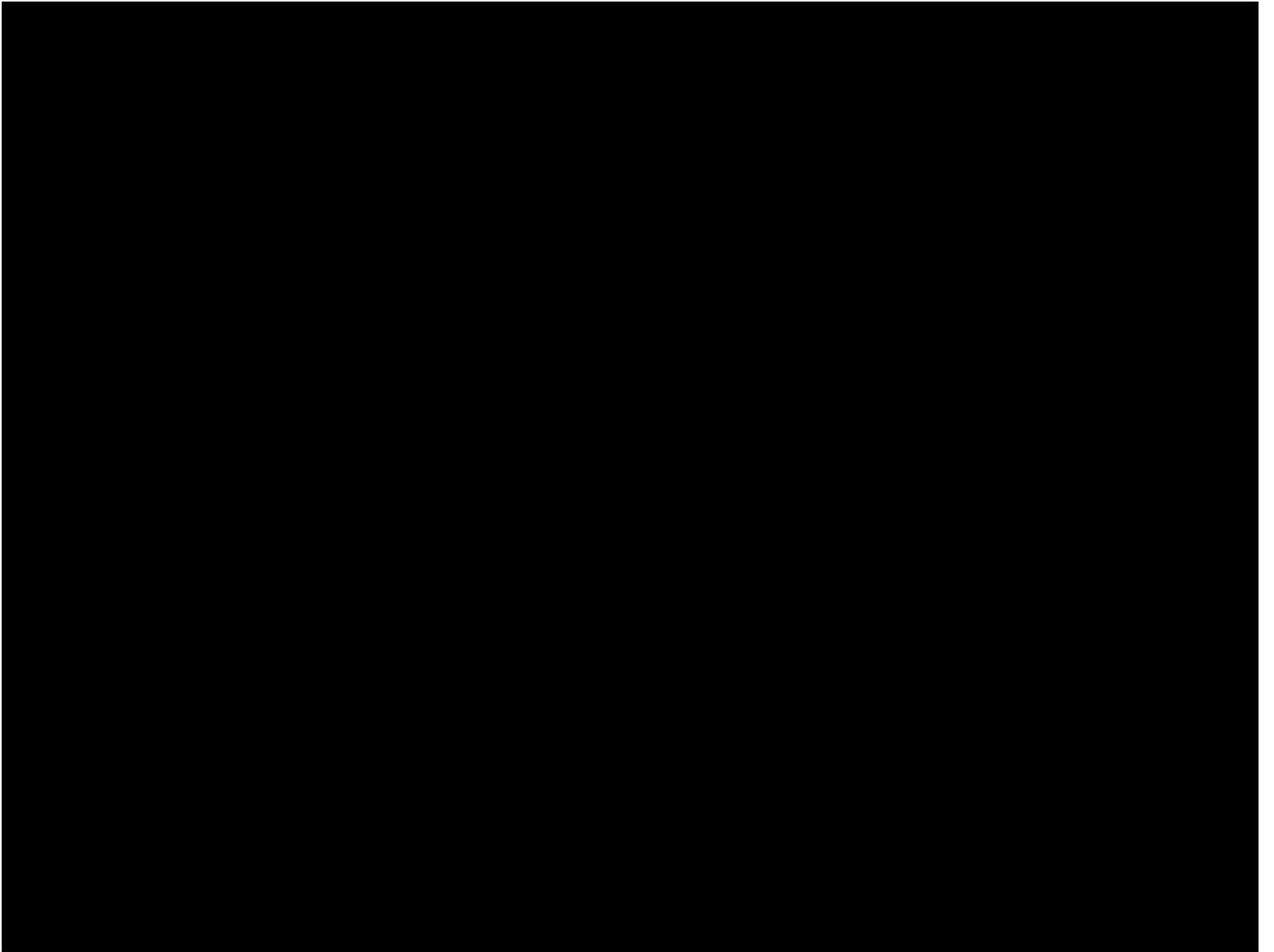
Lewis & Pelham, 1985

Specialized Proteins that Prevent Misfolding and Aggregation - Molecular Chaperones

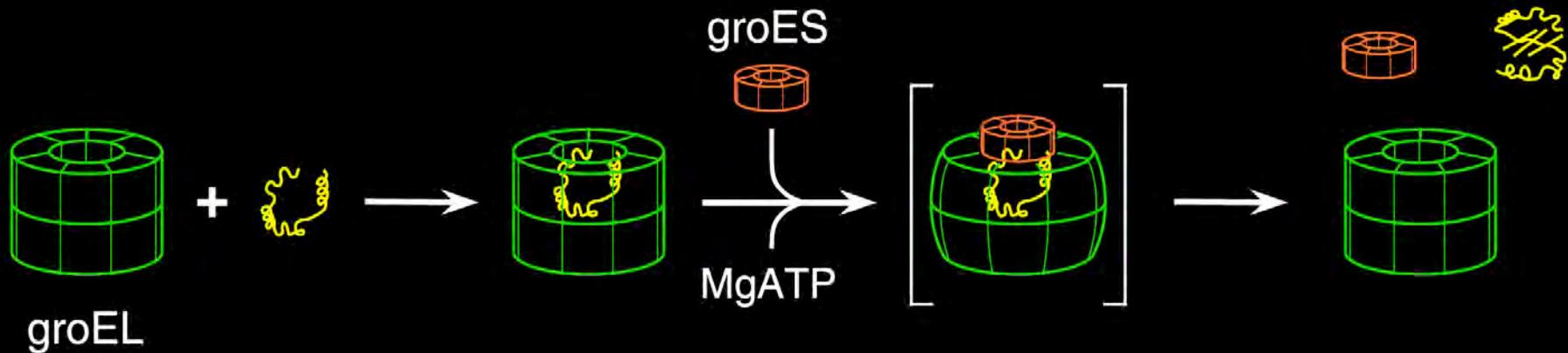
Binding Exposed Hydrophobic Surfaces - Release Mediated by ATP







GroEL-GroES-mediated folding reaction



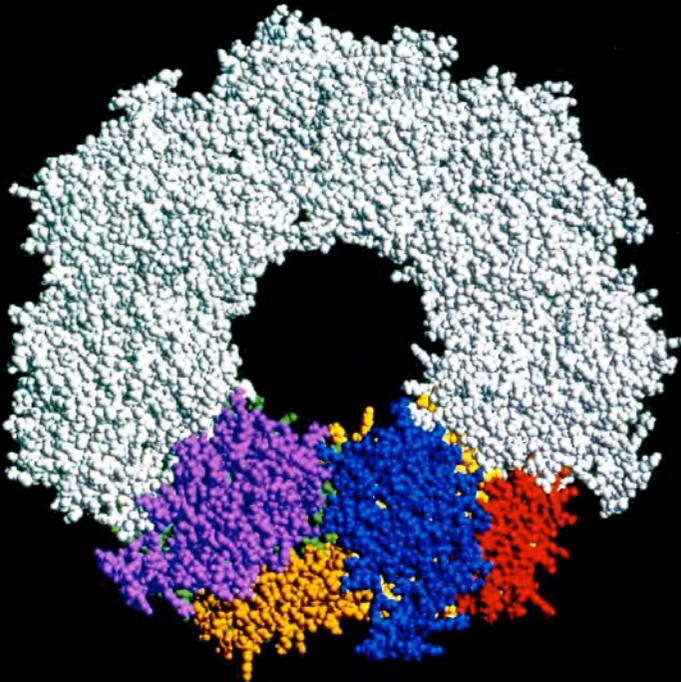
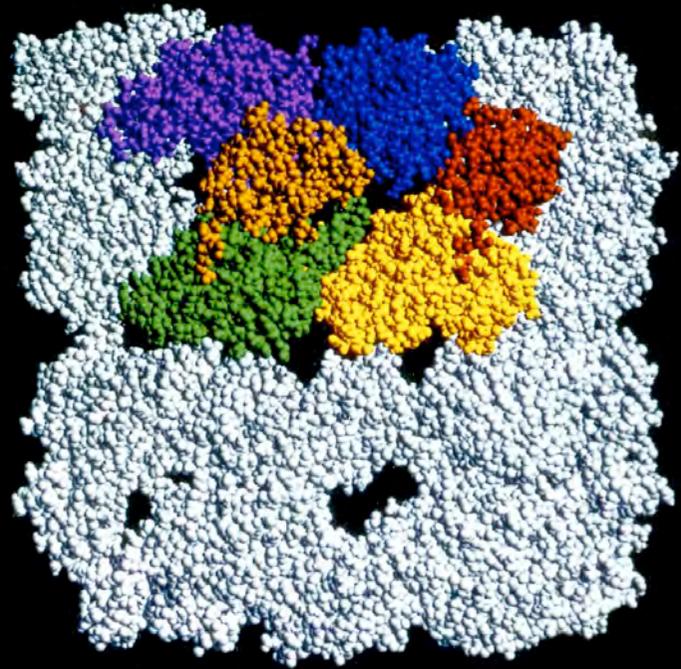
Goloubinoff et al, 1989
Martin et al, 1991

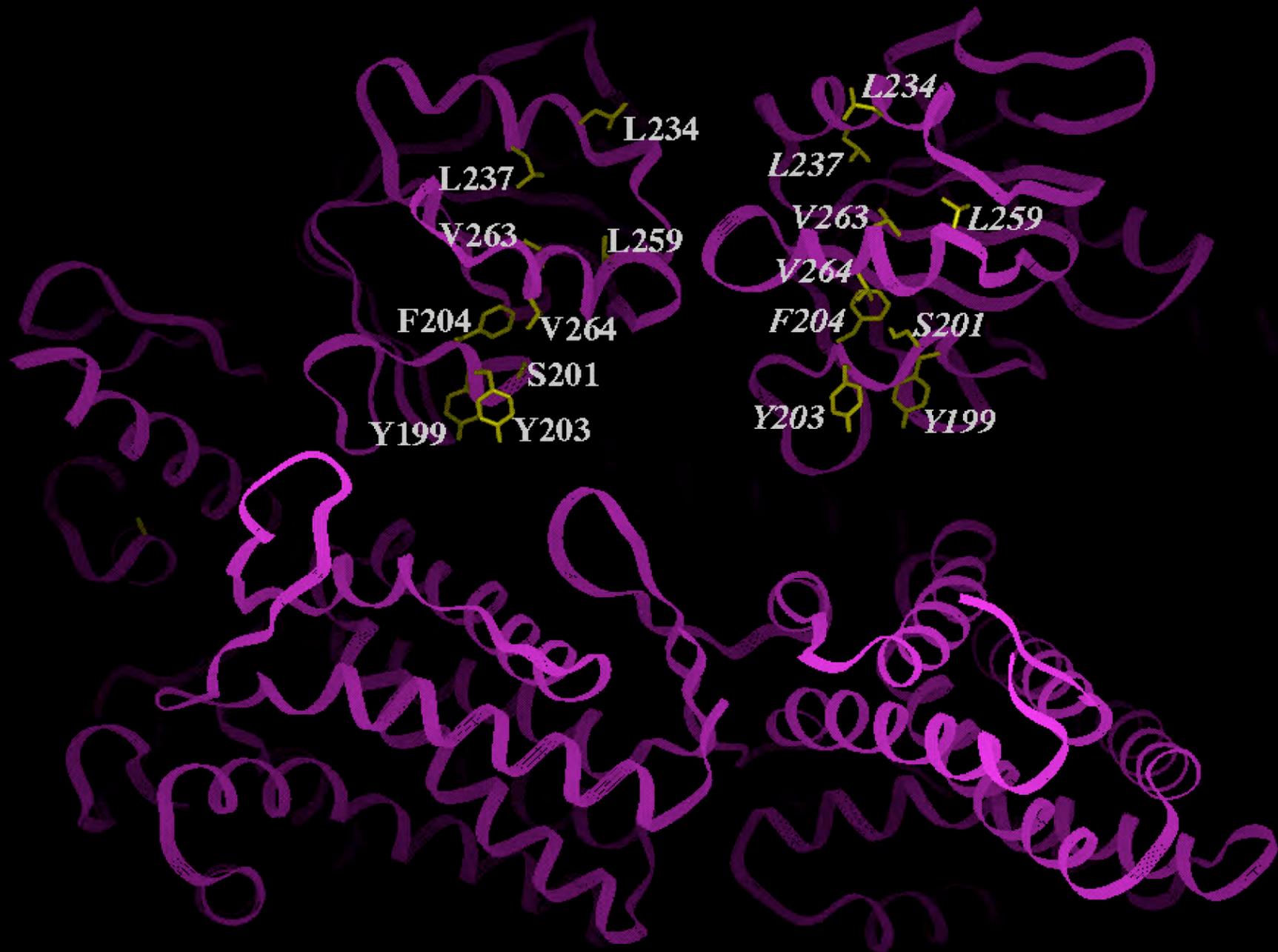
NTP consumption for production of the enzyme rhodanese:

Translation at the ribosome - 4 ~P per peptide bond x 297 aa = 1188 ~P consumed

GroEL/GroES-mediated folding – ~130 ATP hydrolyzed/rhodanese molecule folded =
~10% amount employed for synthesis;

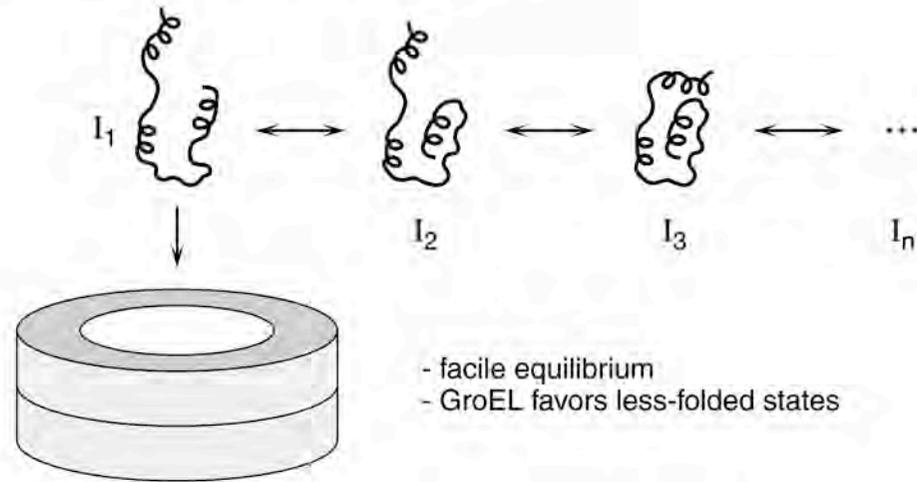
implies multiple turnovers of 7-subunit rings, and amount energy consumed is $\sim 100X \Delta G_{U-N}$



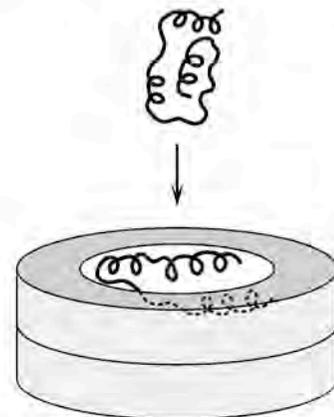


POTENTIAL ACTIONS OF BINDING IN RESCUING MISFOLDED STATES (PULLING NON-NATIVE PROTEINS UP THE ENERGY LANDSCAPE)

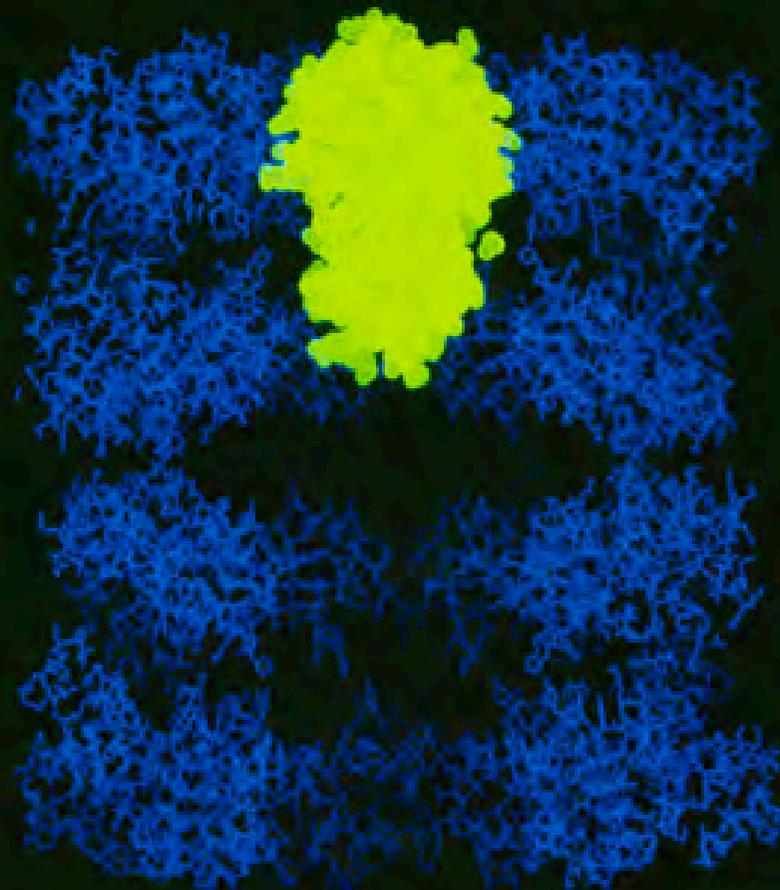
Thermodynamic Partitioning

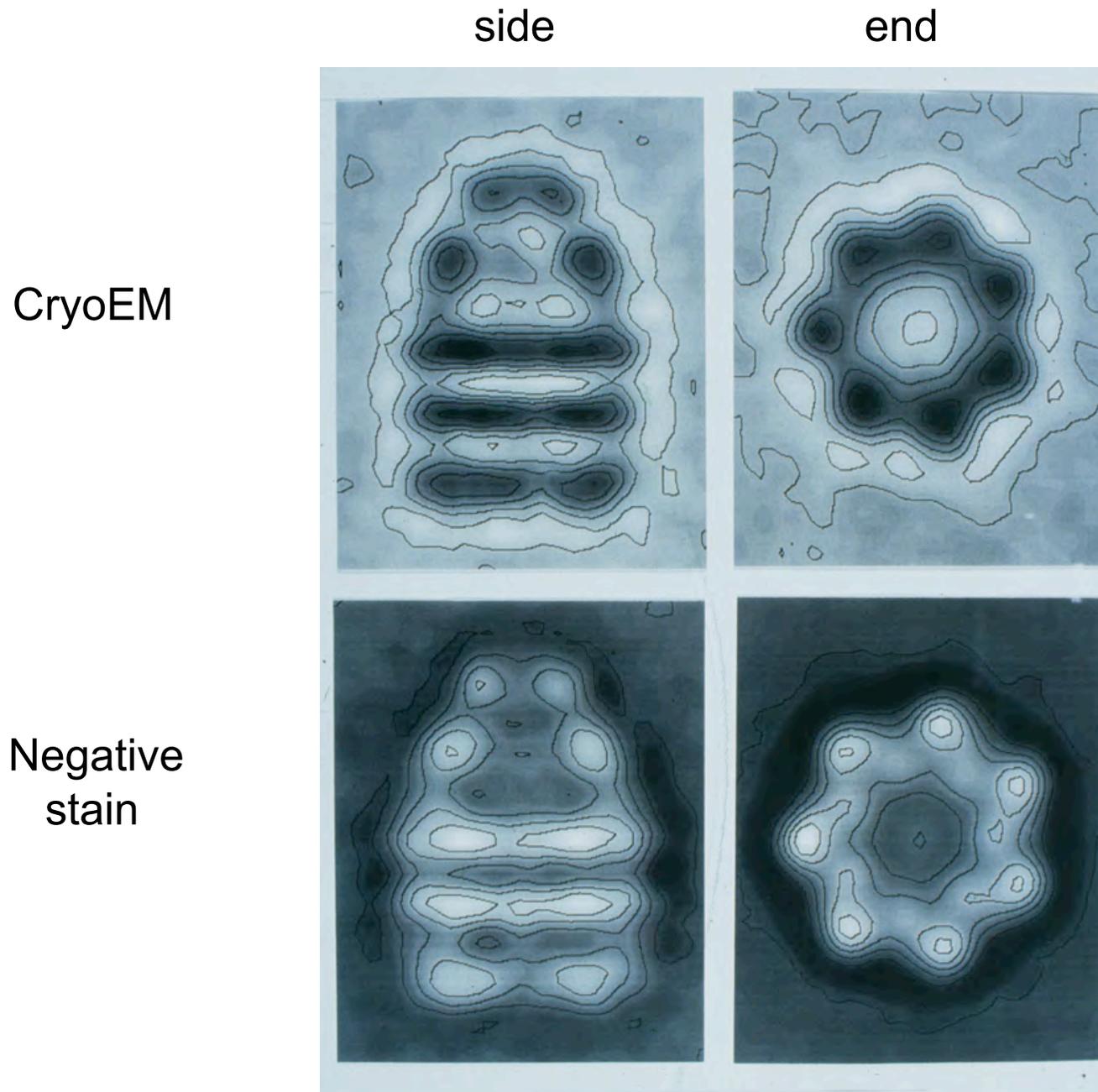


Catalyzed Unfolding



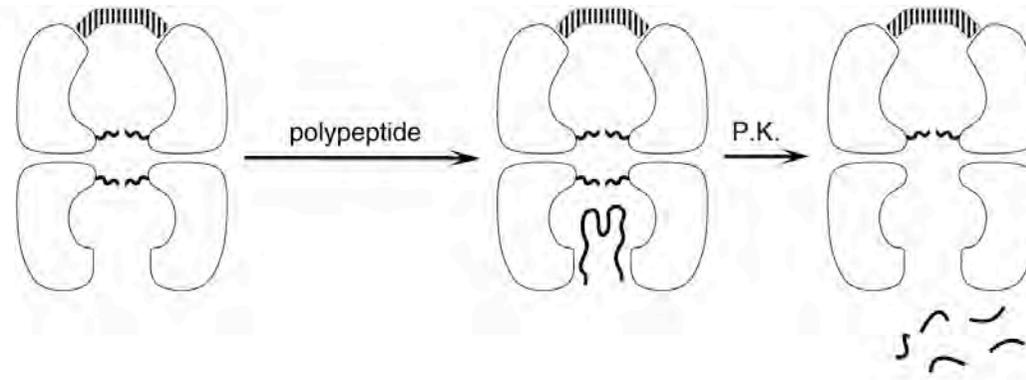
Zahn and Pluckthun, 1994; Walter et al, 1996
Zahn et al, 1996



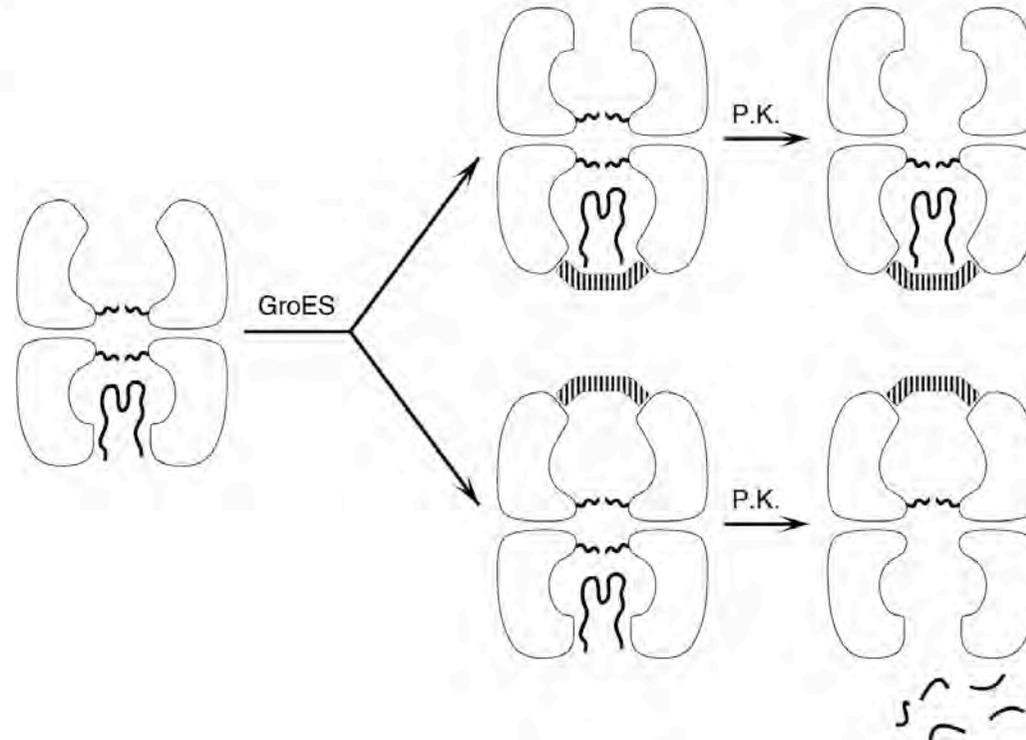


GroEL-GroES-ATP

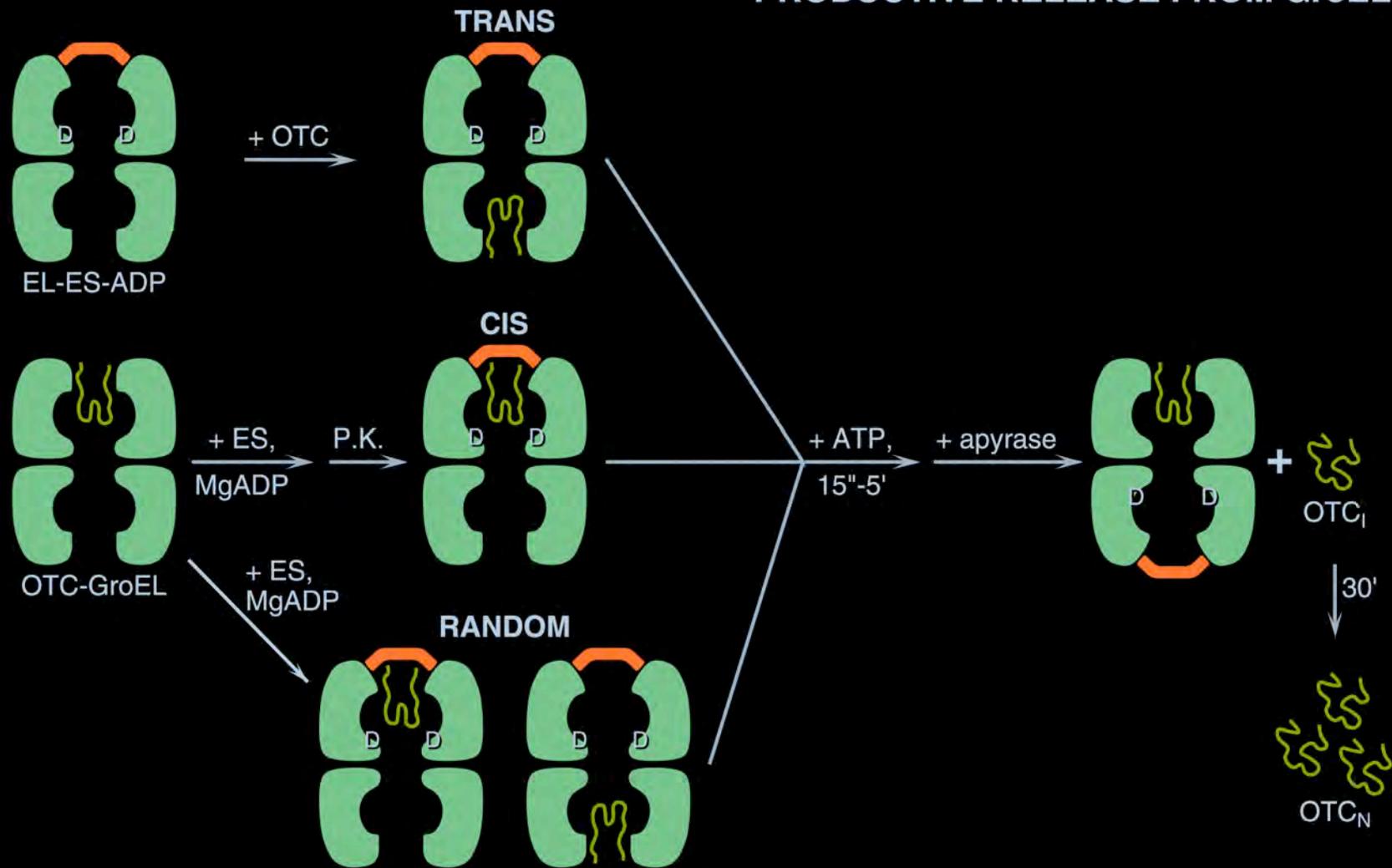
GroES → Polypeptide



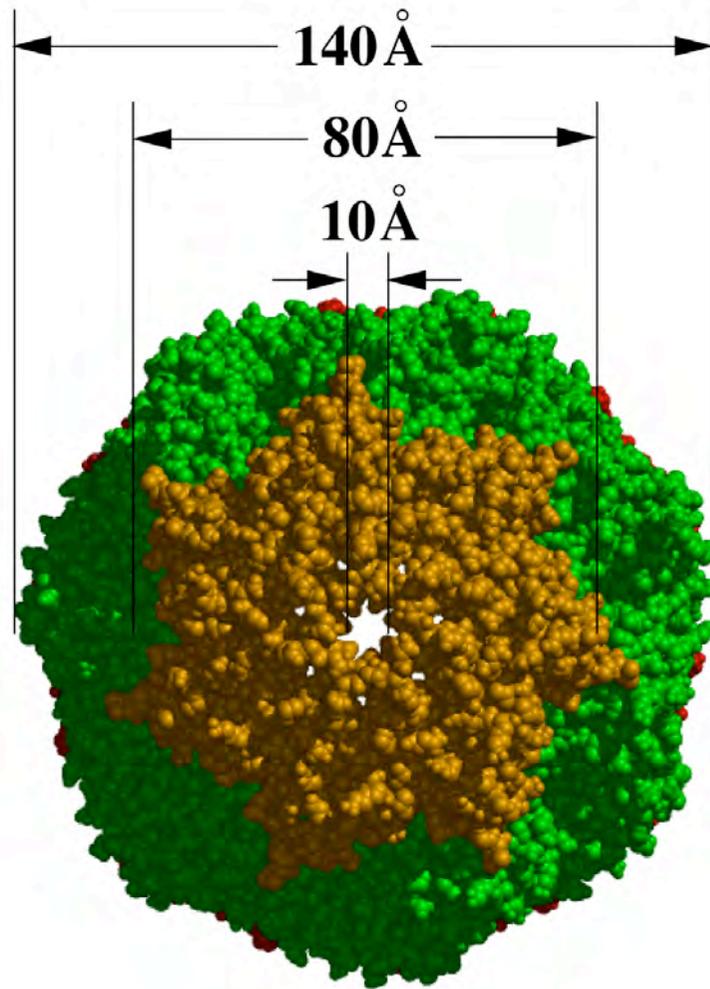
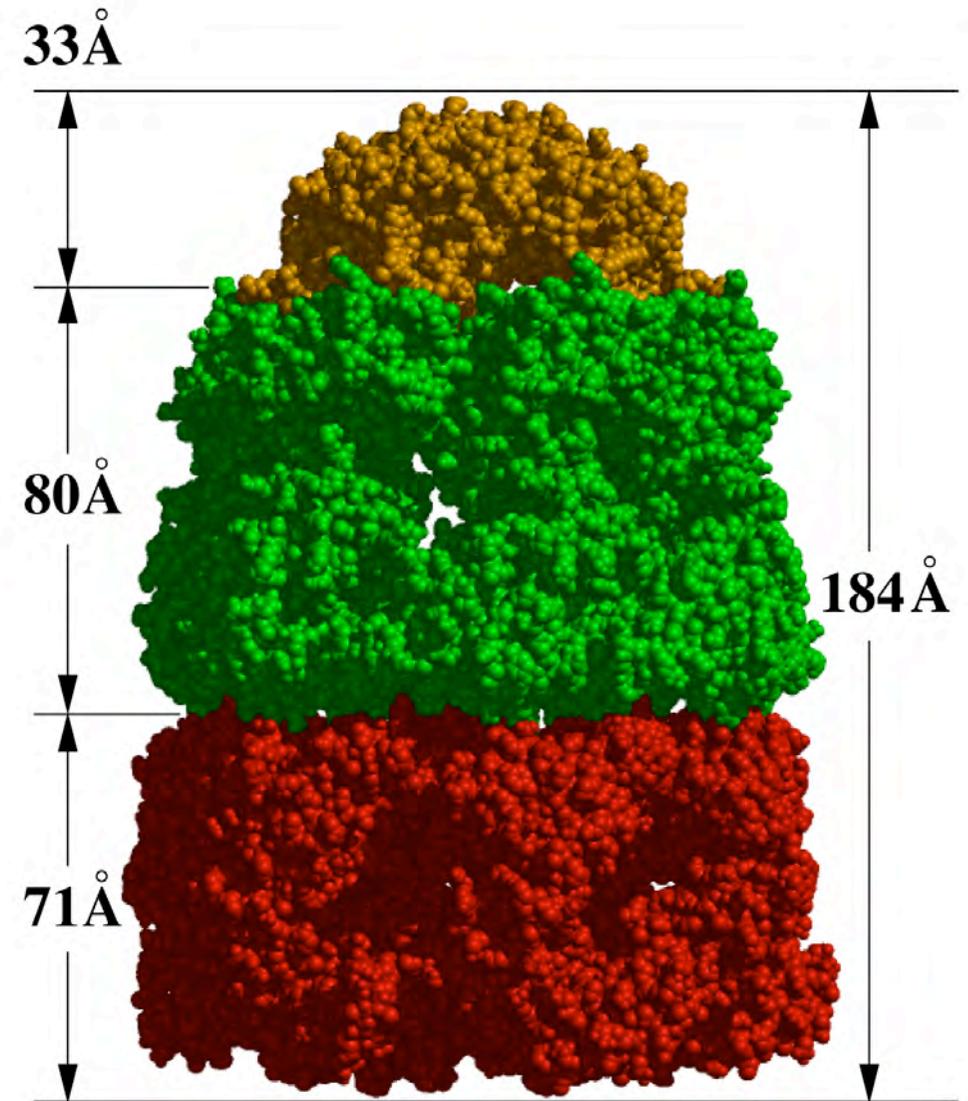
Polypeptide → GroES

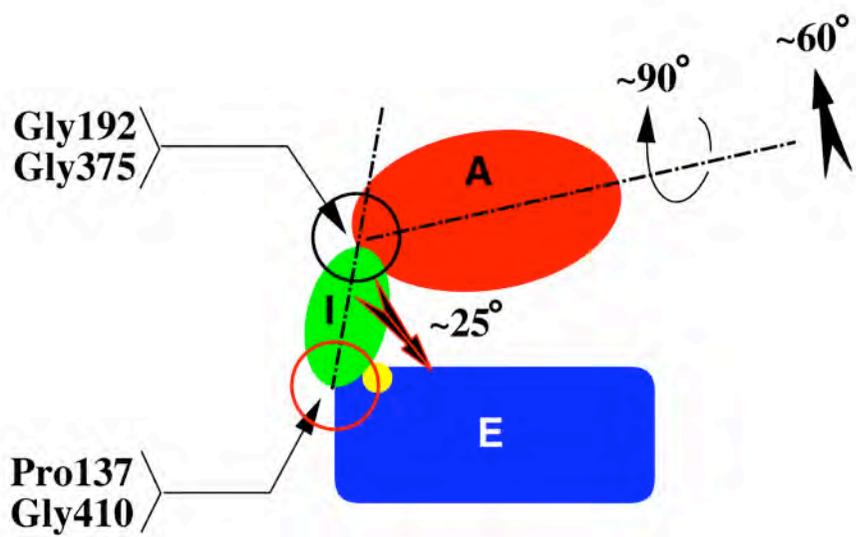
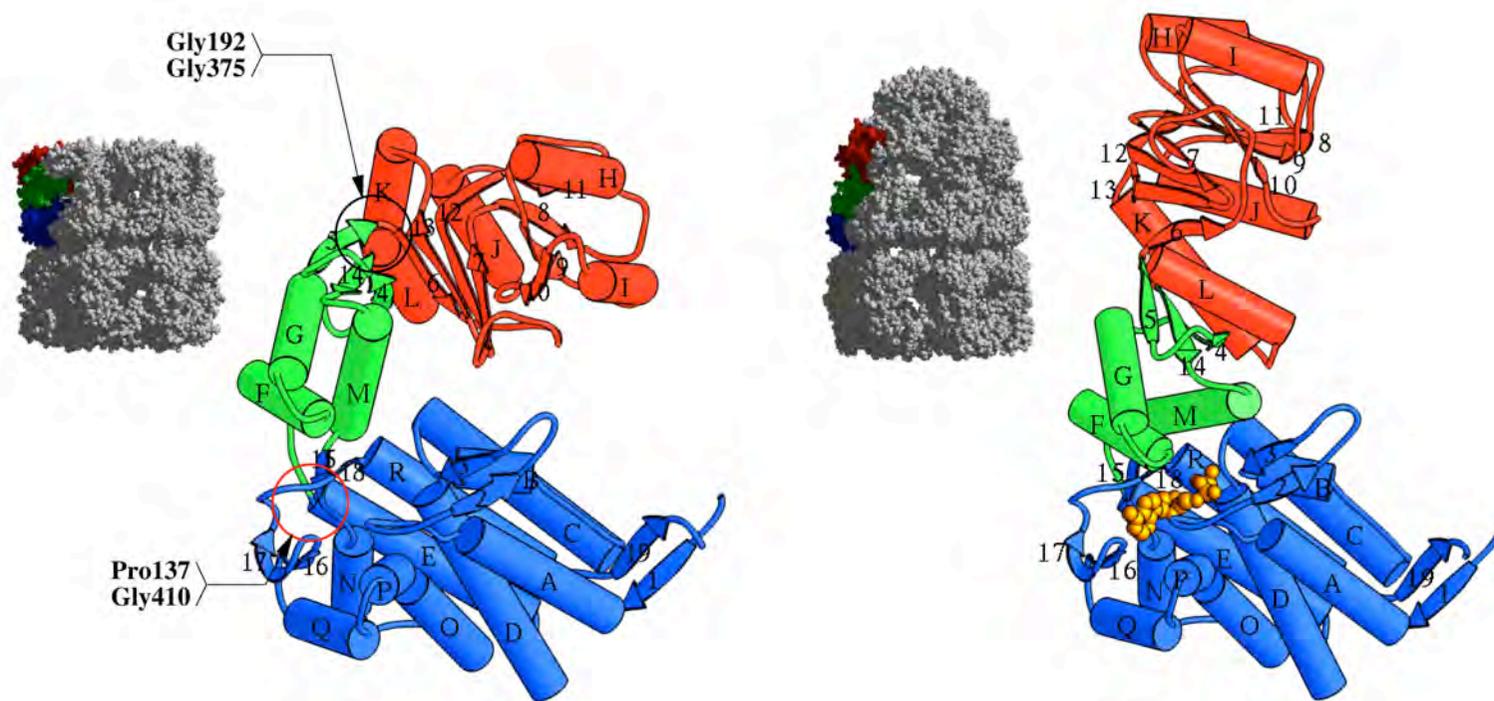


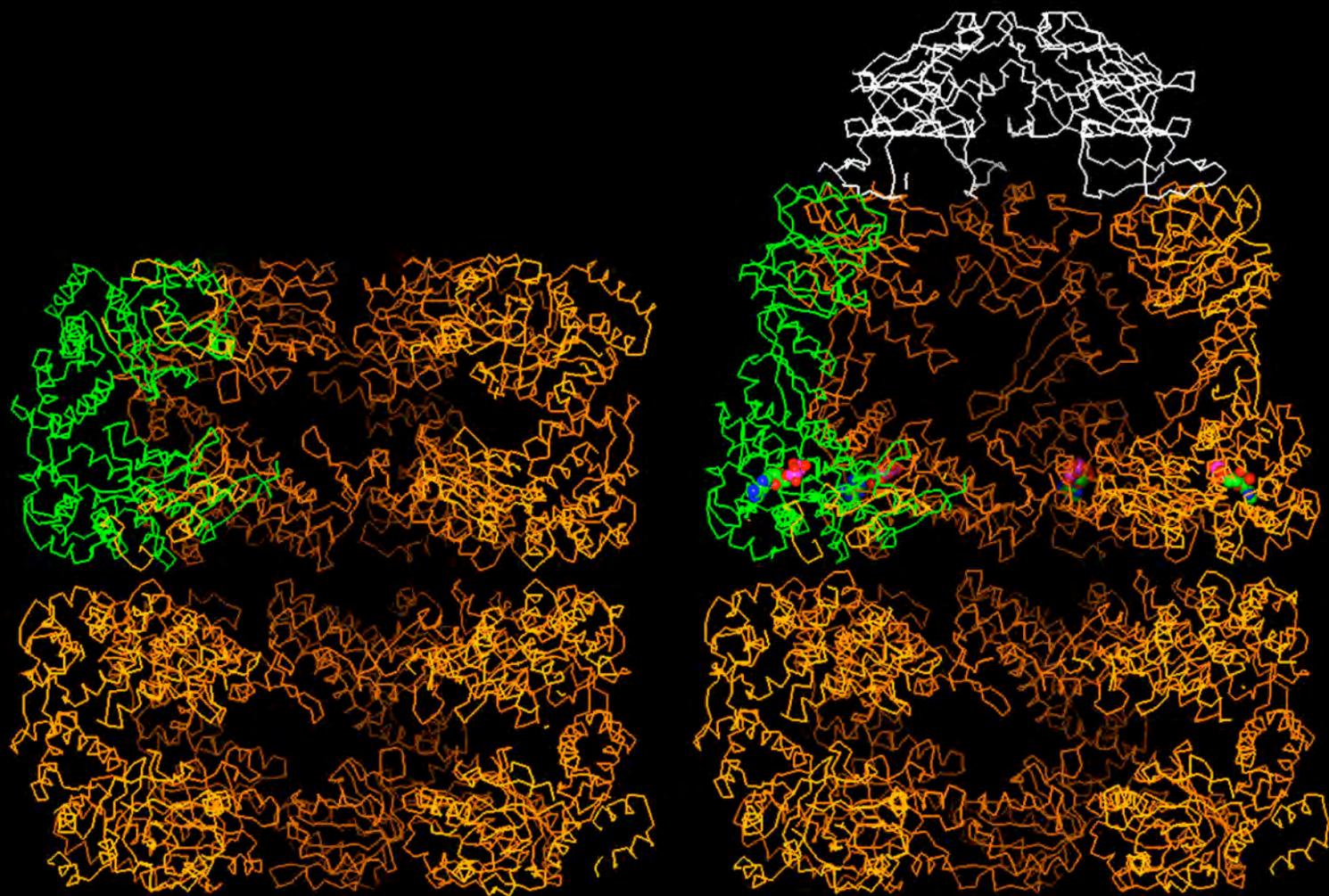
PRODUCTIVE RELEASE FROM GroEL

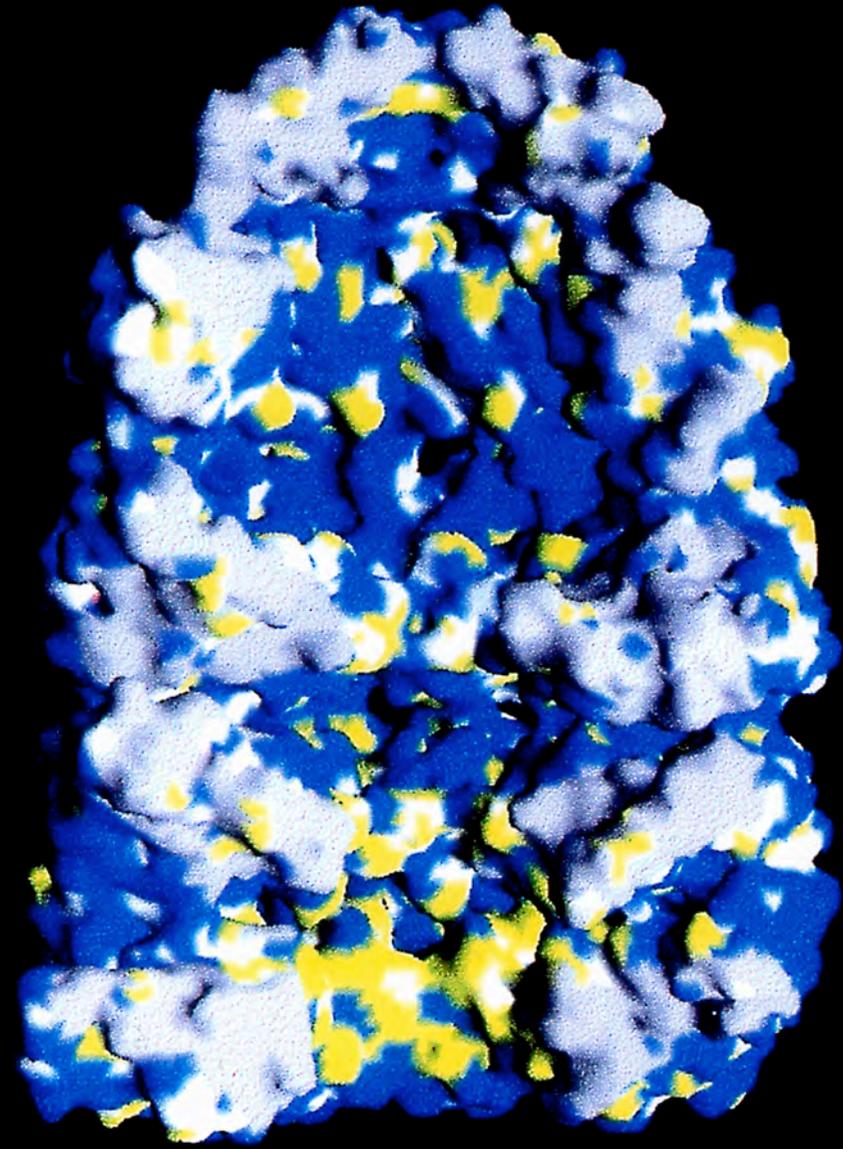




A**B**

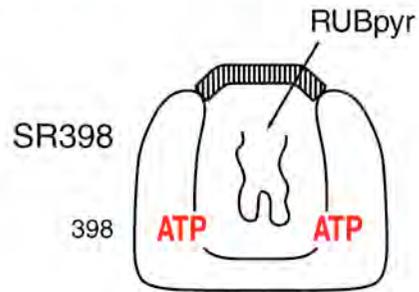






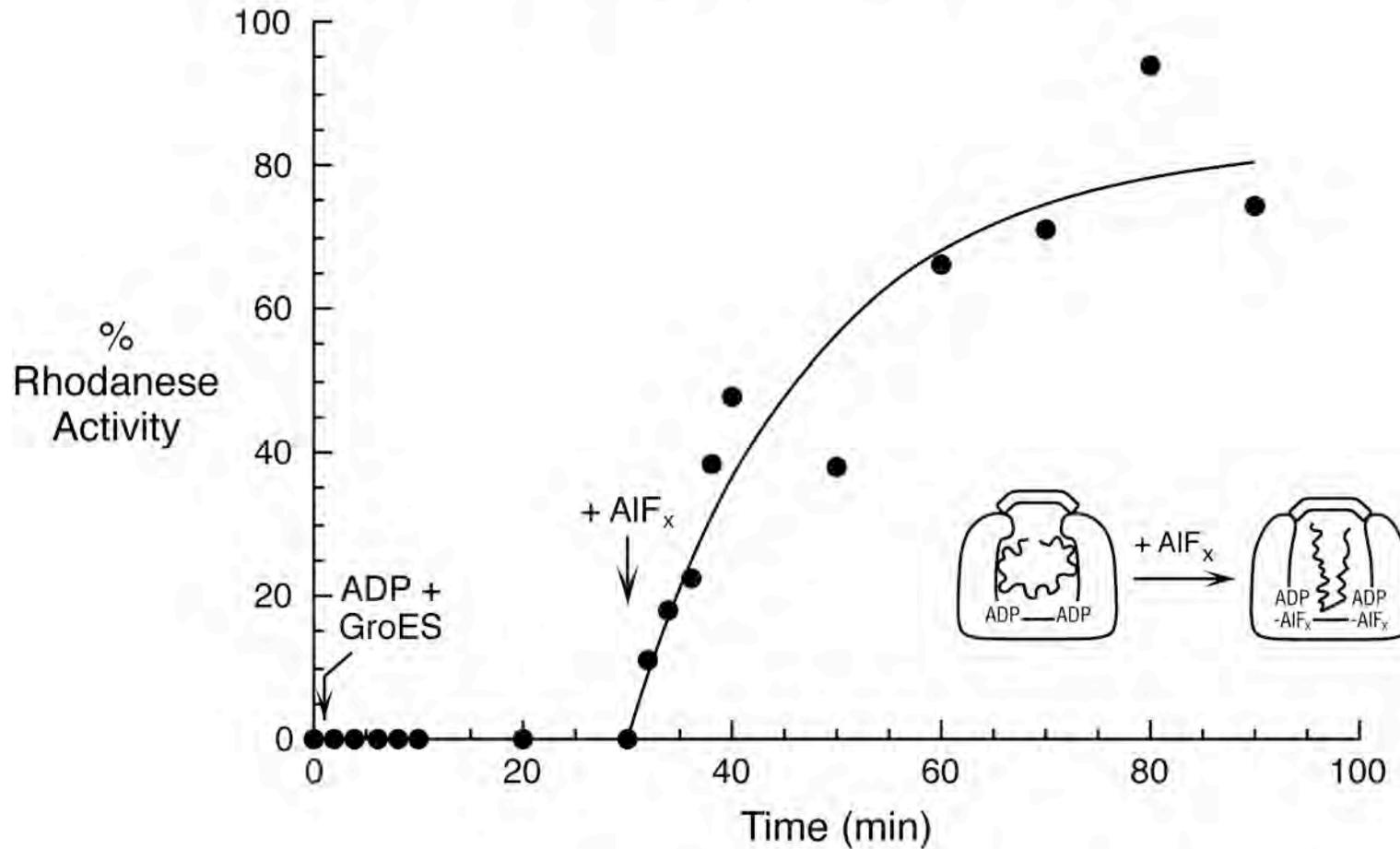
ATP binding, not hydrolysis, triggers productive folding

SR398-GroES-ATP is folding-active

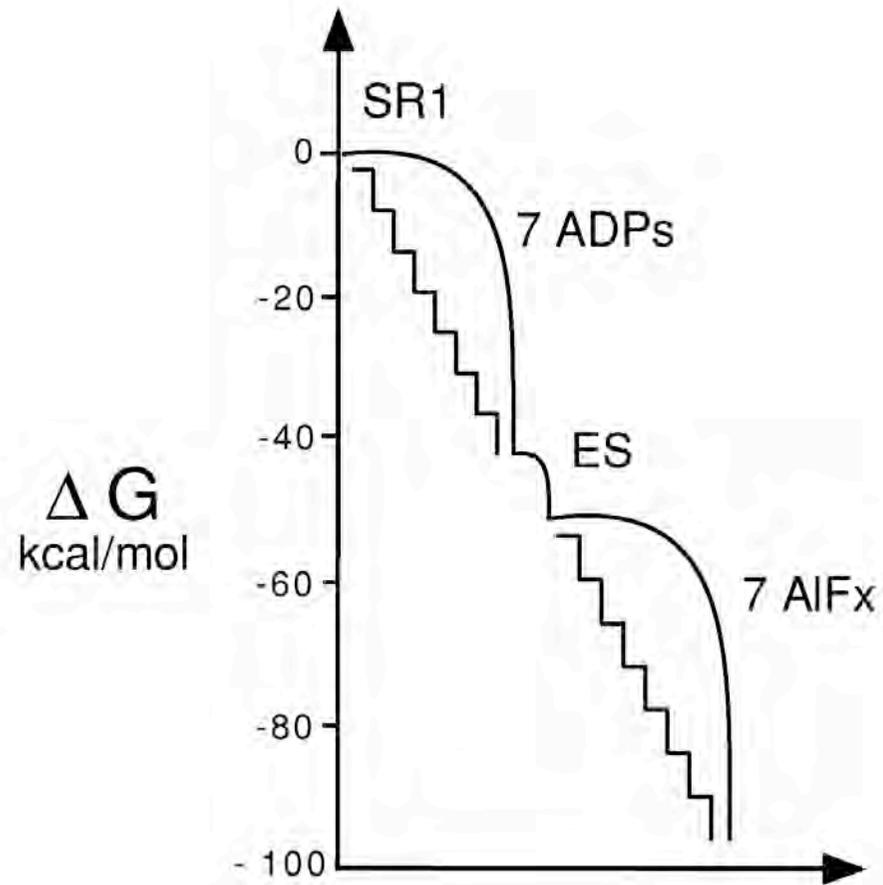


The γ -phosphate of ATP plays an essential role in triggering folding

AlF_x Addition to Preformed SR1-GroES-ADP-Rhodanese Complex Triggers Productive Folding



Estimated free energy transitions during activation of SR1

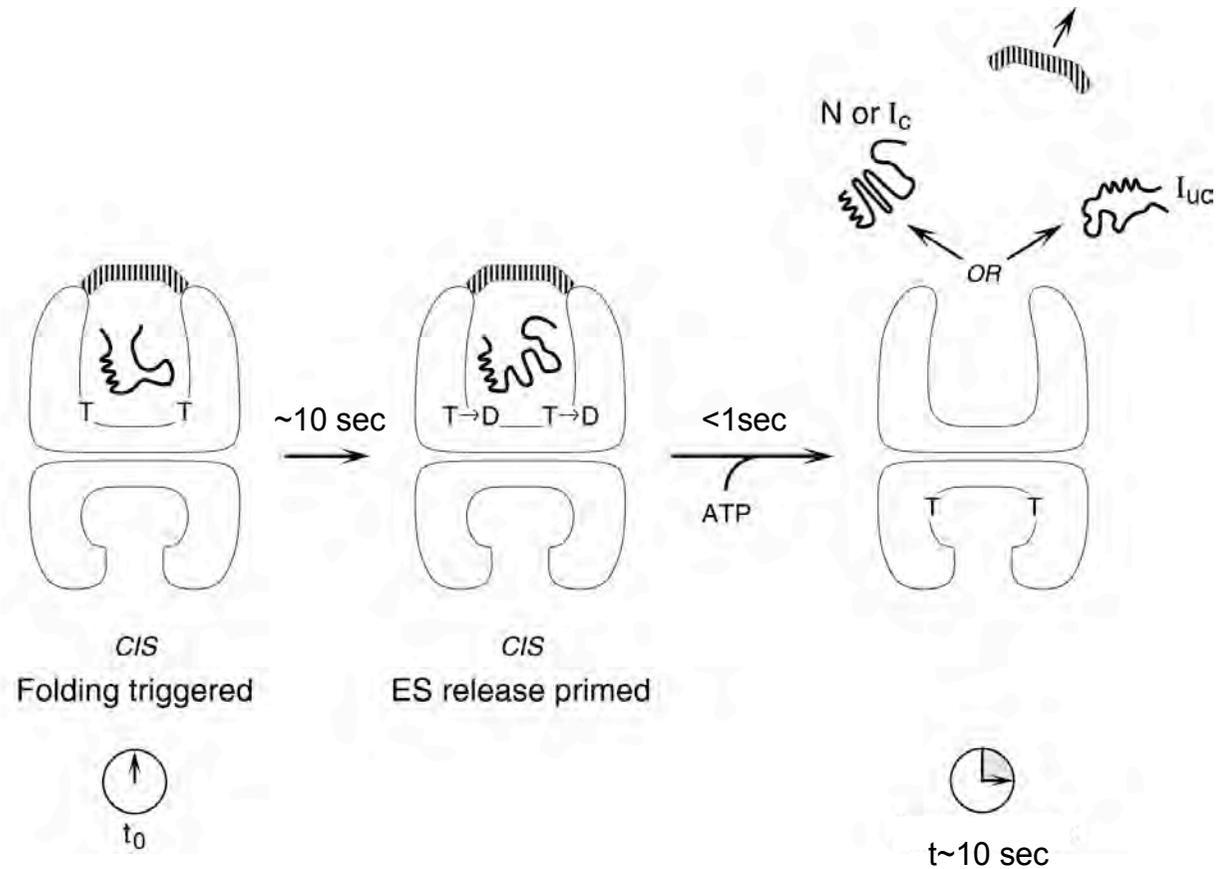


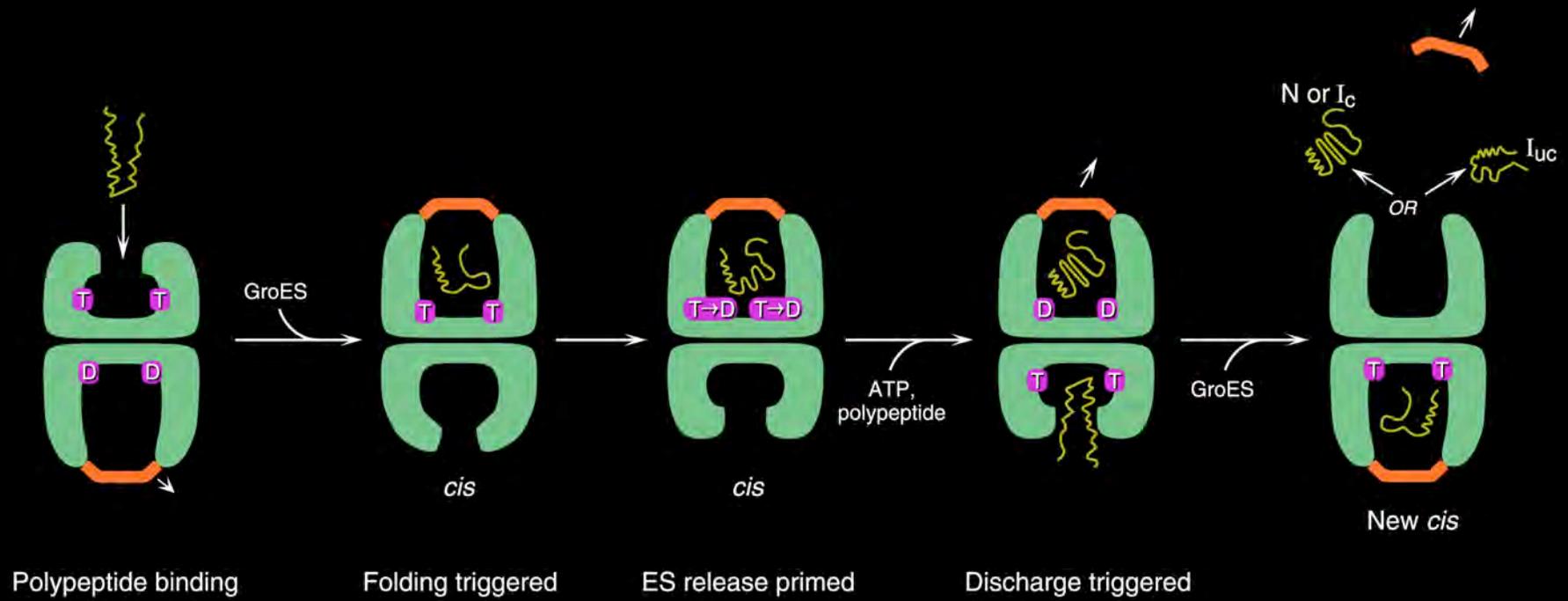
ADP binding $K_D = 30 \mu\text{M}$

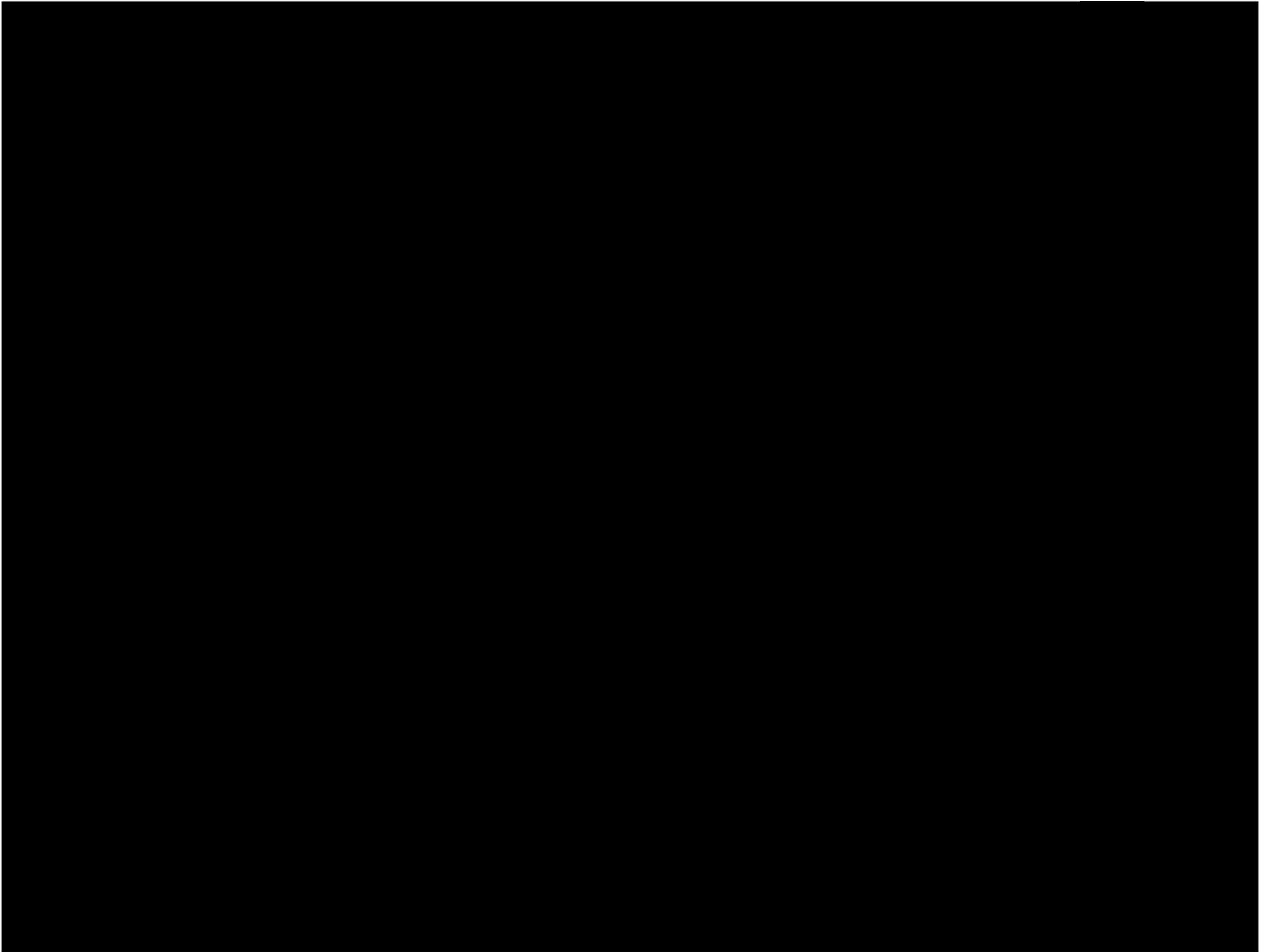
GroES binding $K_D = 0.45 \mu\text{M}$

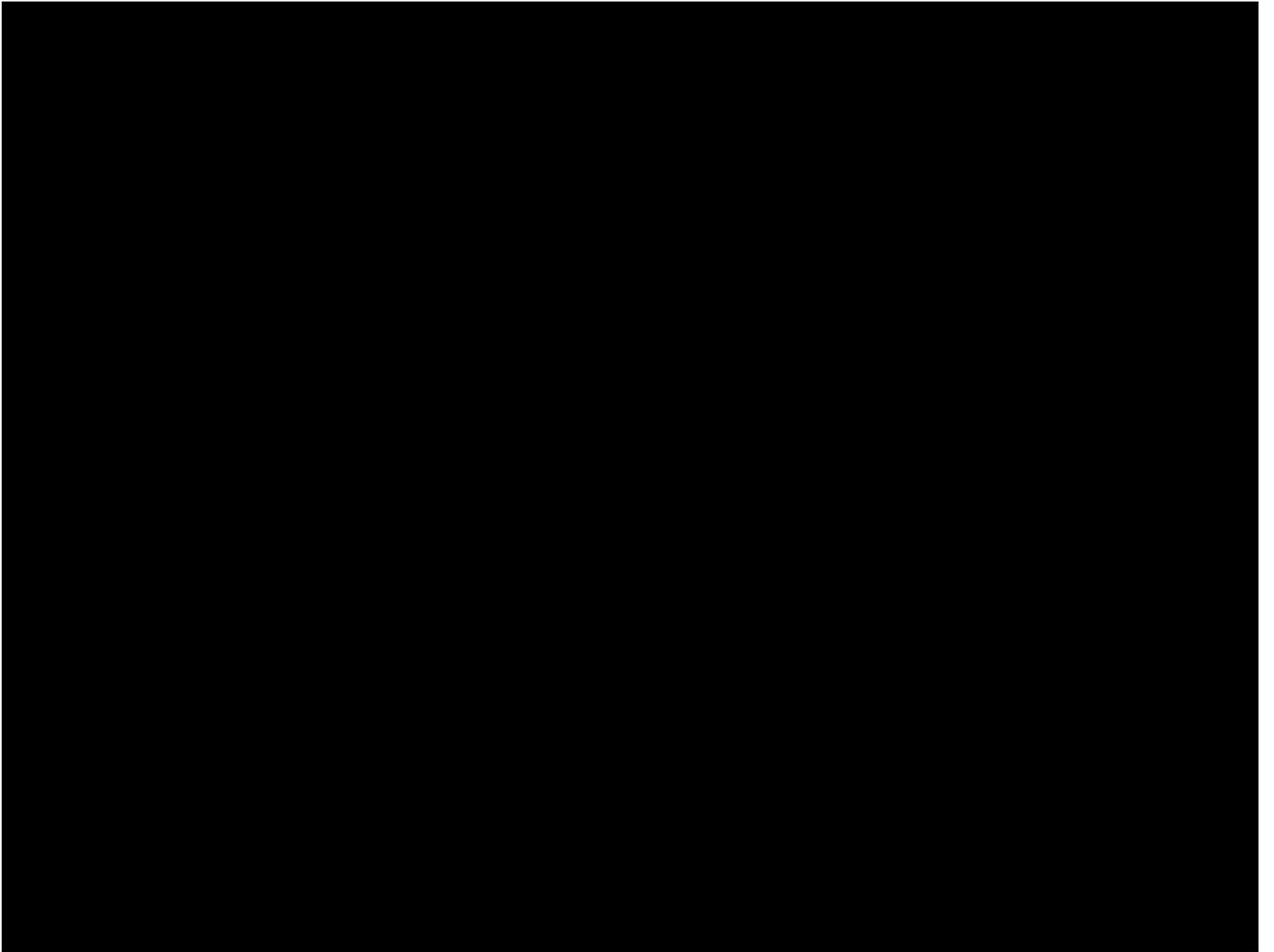
AIFx binding $K_D = 16 \mu\text{M}$

ATP hydrolysis at GroEL serves to advance the reaction forward, acting as a “timer”









Hsp60/GroEL

Ming Cheng
Ulrich Hartl (Munich)

X-ray and EM work

Kerstin Braig
Charu Chaudhry
Zbyszek Otwinowski
Zhaouhui Xu
Helen Saibil (Birkbeck)
Paul Sigler

Topology and
ATP cycle

George Farr
Wayne Fenton
Hays Rye
Jonathan Weissman

NMR studies

Eric Bertelsen
Jocelyne Fiaux
Reto Horst (Scripps)
Peter Wright (Scripps)
Kurt Wüthrich (Scripps)

